# Machine Learning in Structure Biology

Yang Zhang

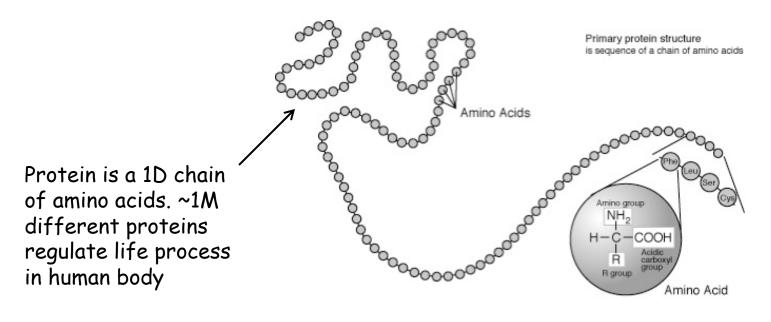
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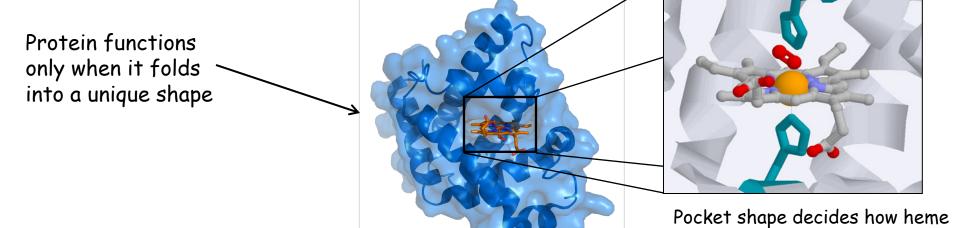
University of Michigan

### <u>Case Studies of Machine-Learning in</u> <u>Structure Biology</u>

- 1. Protein Secondary Structure Prediction
- 2. Protein Contact Prediction
- 3. Disease-Associated Mutation Prediction

### What is protein?





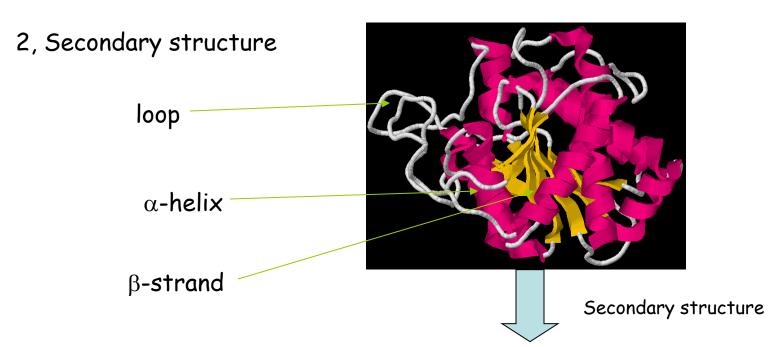
3D structure of myoglobin

binds and how oxygen is transferred through blood

### 1.1. What is protein secondary structure?

1, Primary amino acid sequences (1D)

MVLSEGEWQLVLHVWAKVEADVAGHGQDILIRLFKSHPETLEKFDRV KHLKTEAEMKASEDLKKHGVTVLTALGAILKKKGHHEAELKPLAQSHA TKHKIPIKYLEFISEAIIHVLHSRHPGNFGADAQGAMNKALELFRKDI AAKYKELGYQG

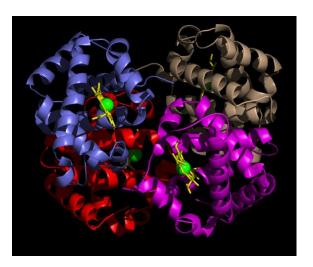


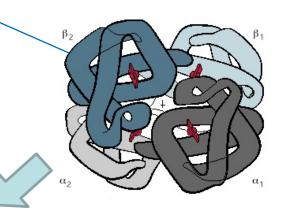
### 1.1. What is protein secondary structure?

### 3, Tertiary structure



4, Quaternary structure

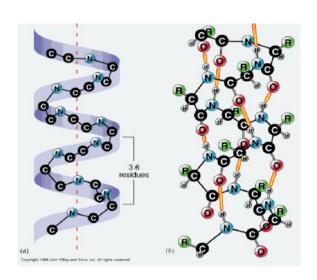


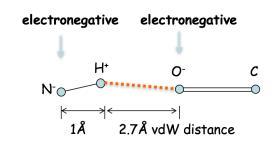


### 1.2. Hydrogen-bond

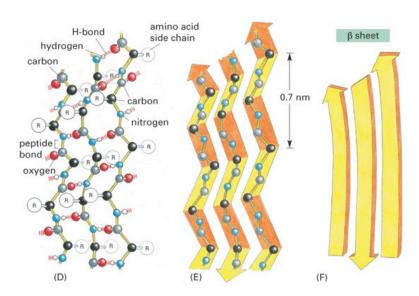
(Secondary structure is specified by H-bonding)

### H-bond in $\alpha$ -helix





### H-bond in $\beta$ -sheet

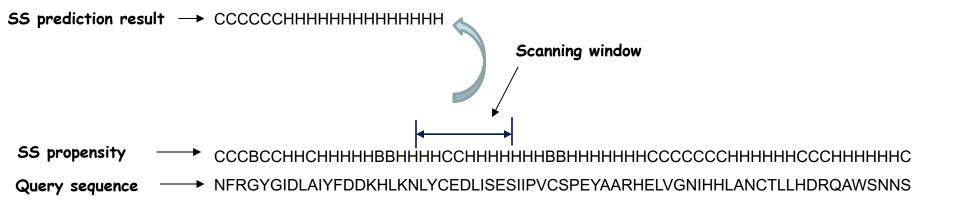


## 1.3. How to predict second structure from seuqence? (Former effort: Chou-Fasman method)

### SS propensity of amino acids:

Helical Residues	$P_{\alpha}$	β-Sheet Residues <sup>c</sup>	$P_{\beta}$	
Glu <sup>(-)</sup>	1.53)	Met	1.67	
Ala	$1.45 H_{\alpha}$	Val	1.65	$H_{\beta}$
Leu	1.34	Ile	1.60	Predicting SS based on simple
His(+)	1.24	Cys	1.30	/ statistics
Met	1.20	Tyr	1.29	
Gln	1.17 h	Phe	1.28	
Trp	$1.17$ $h_{\alpha}$	Gln	1.23	$h_B$
Val	1.14	Leu	1.22	
Phe	1.12)	Thr	1.20	
Lys(+)	1.07	Trp	1.19	
Ile	$1.00$ $I_{\alpha}$	Ala	0.97	$\mathbf{I}_{\mathcal{B}}$
Asp(-)	0.98	Arg(+)	0.90	
Thr	0.82	Gly	0.81	i <sub>s</sub>
Ser	$0.79$ } $i_{\alpha}$	Asp(-)	0.80	
Arg(+)	0.79	Lys(+)	0.74	
Cys	0.77	Ser	0.72	
Asn	0.73	His(+)	0.71	$b_{\beta}$
Tyr	$0.61$ $b_{\alpha}$	Asn	0.65	
Pro	0.50)	Pro	0.62	
Gly	$0.53$ $\mathbf{B}_{\alpha}$	Glu <sup>(-)</sup>	0.26}	$B_{\beta}$

### Chou-Fasman method



### Accuracy of SS prediction:

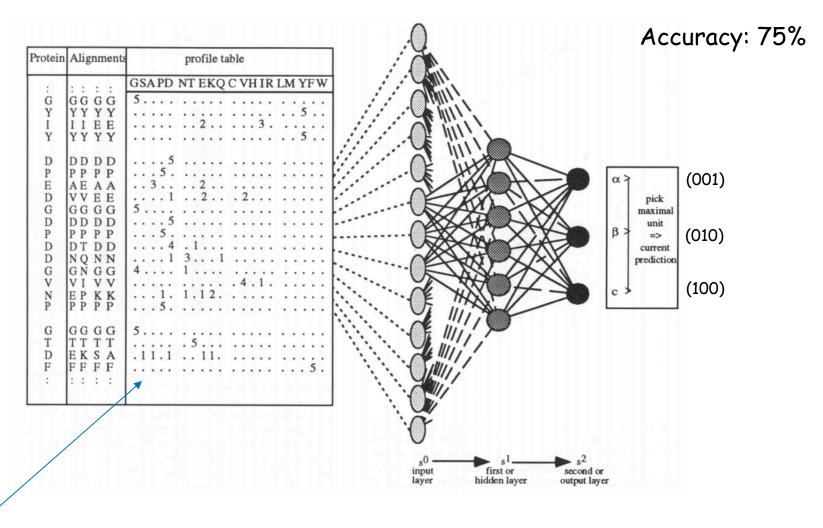
$$Q3 = \frac{\#residues\ with\ correctled\ predicted\ SS}{\#total\ residues}$$

Average accuracy: 50-60%

Better than random: 47%

(32%  $\alpha$ -helix, 21%  $\beta$ -strand, 47% loop)

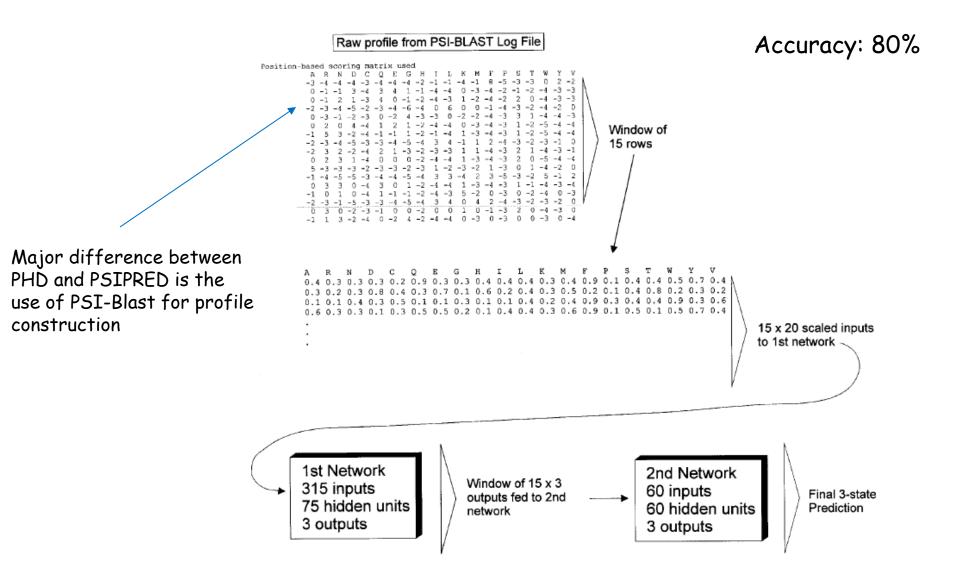
### 1.3. Former effort on SSP: PHD



Using sequence profile instead of single sequence as input of network training

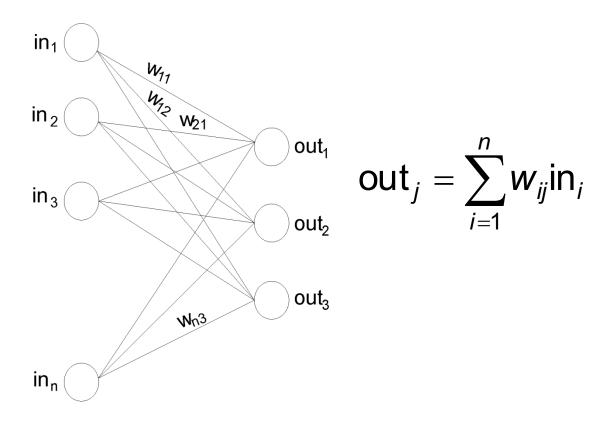
Rost, B. & Sander, C., Prediction of protein secondary structure at better than 70 % Accuracy, Journal of Molecular Biology, (1993) 232, 584-599.

### 1.3. Former effort on SSP: PSIPRED



Jones, D., Protein secondary structure prediction based on position-specific scoring matrices. J. Mol. Biol, (1999) 292, 195-202.

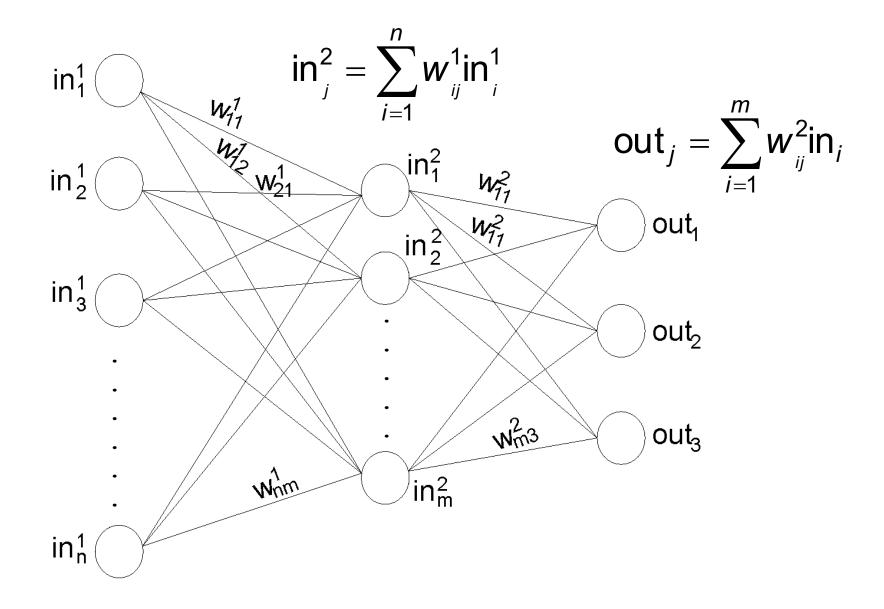
### What is neural network?



The principle of neural network is to adjust the weights  $(w_{ij})$  iteratively so that output  $(out_j)$  is close to the true answer (T).

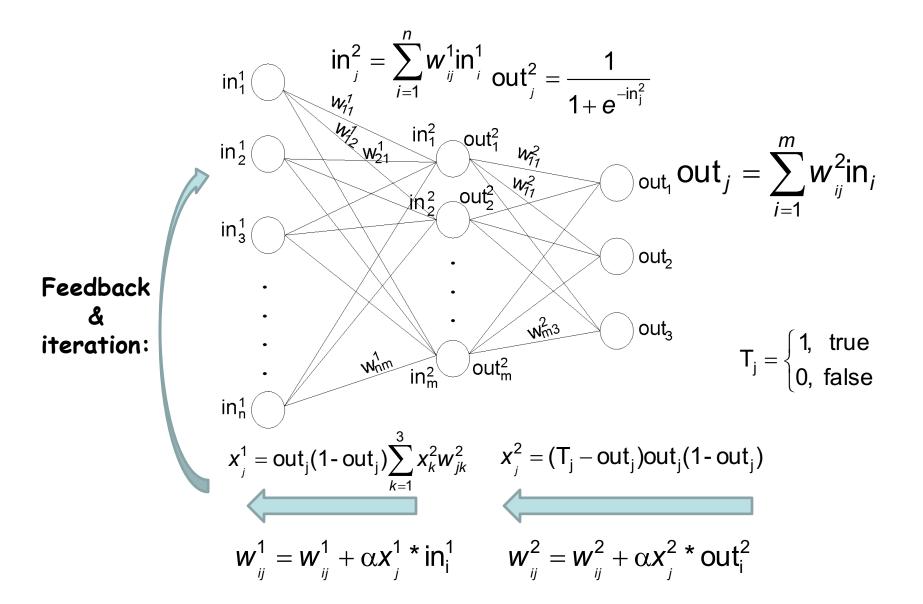
$$w_{ij}^{new} = w_{ij}^{old} + \alpha f(error)$$

### A two layer network

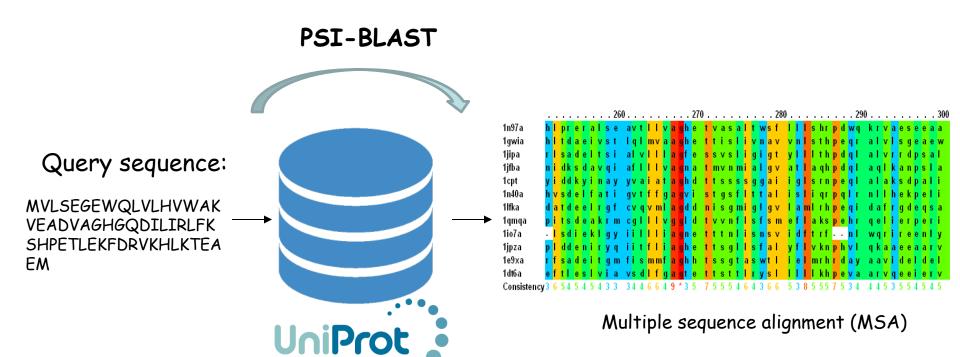


### 1.4. State of the art: PSSpred

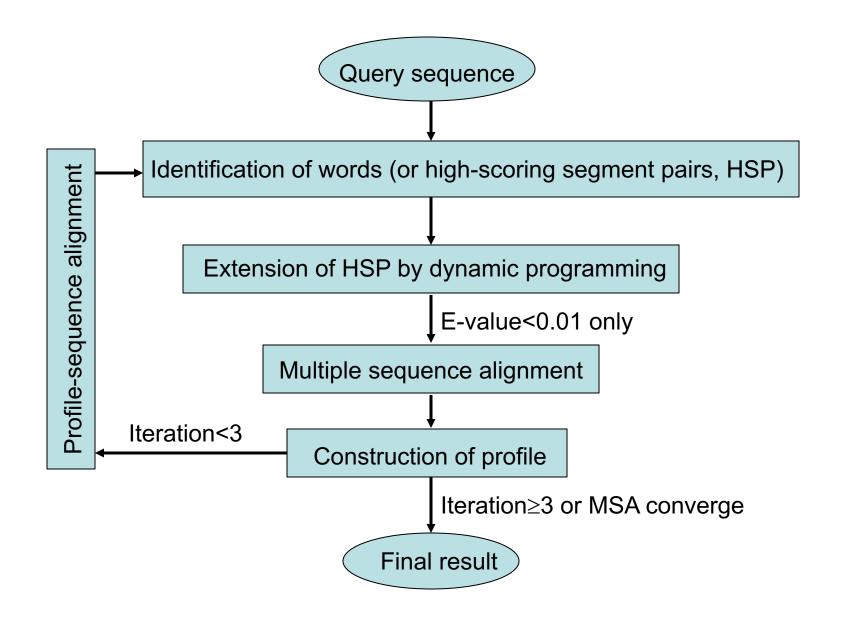
(http://zhanglab.ccmb.med.umich.edu/PSSpred)



### 1.4. PSSpred feature collection



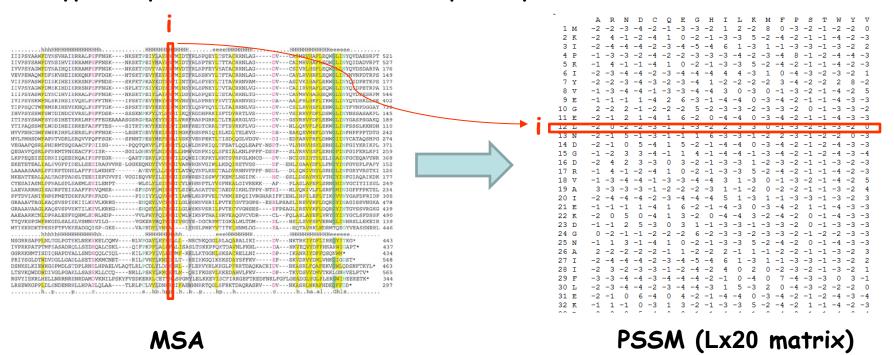
### PSI-Blast Pipeline: Iterative Profile-sequence Alignment Algorithm



5. F. Altschul et al, Nucleic Acids Res. 1997, 25(17):3389-402.

### 1.4. PSSpred feature collection

Five types of profiles are derived from sequence profile

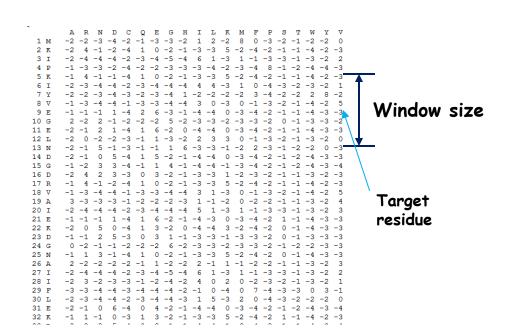


Five profiles derived from PSI-Blast MSA 
$$\begin{aligned} & in = 1/(1+e^{-x}) & \text{x=log}(Q_{ij}/P_i) \\ & \text{MTX:} & in = 1/(1+e^{-x/100}) & \text{x=log}(Q_{ij}/P_i) \\ & \text{PROF}_{W} : & in = 1/(1+e^{-x}) & \text{x=} \sum_{a=1}^{20} \sum_{k=1}^{f(a,j)} w(k)B(A_i,a) \\ & \text{FREQ}_{CW} : & in = 1/[1+e^{-25(x-\langle x \rangle)}] & \text{x=} \sum_{k=1}^{f(A,j)} w(k) \\ & \text{FREQ}_{CWQ} : & in = 1/[1+e^{-30(x-\langle x \rangle)}] & \text{x=} \sum_{k=1}^{f(A,j)} w(k) \end{aligned}$$

### 1.4. PSSpred training parameters

### Seven PPSpred programs

- 1. mtx\_pssm\_freqccw\_profw\_12
- mtx\_freqccw\_profw\_freqccwG\_15
- mtx\_freqccw\_profw\_freqccwG\_12
- 4. mtx\_freqccw\_profw\_12
- 5. mtx freqccw profw 18
- 6. mtx\_profw\_freqccwG\_18
- 7. mtx\_profw\_12

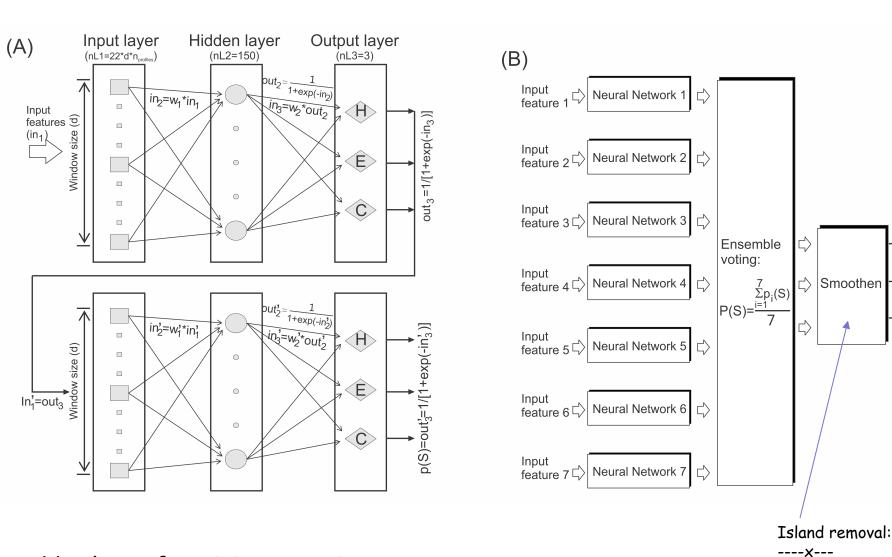


Predictors	Training	Windo	Number of
	features	w size	iterations
PSSpred1	PSSM	12	44
	MTX		
	PROF W		
	FREQ CW		
PSSpred2	MTX	15	38
	PROF W		
	FREQ CW		
	FREQ CWG		
PSSpred3	MTX	12	62
i oopieus	PROF W	12	02
	FREQ CW		
	FREQ CWG		
	_		
PSSpred4	MTX	12	63
	PROF_W		
	FREQ_CW		
PSSpred5	MTX	18	54
	PROF_W		
	FREQ_CW		
PSSpred6	MTX	18	47
	PROF_W		
	FREQ_CWG		
PSSpred7	MTX	12	84
•	PROF W		

### 1.4. Pipeline of PSSpred

 $\langle \hat{H} \rangle$ 

---XX---



### Number of training proteins:

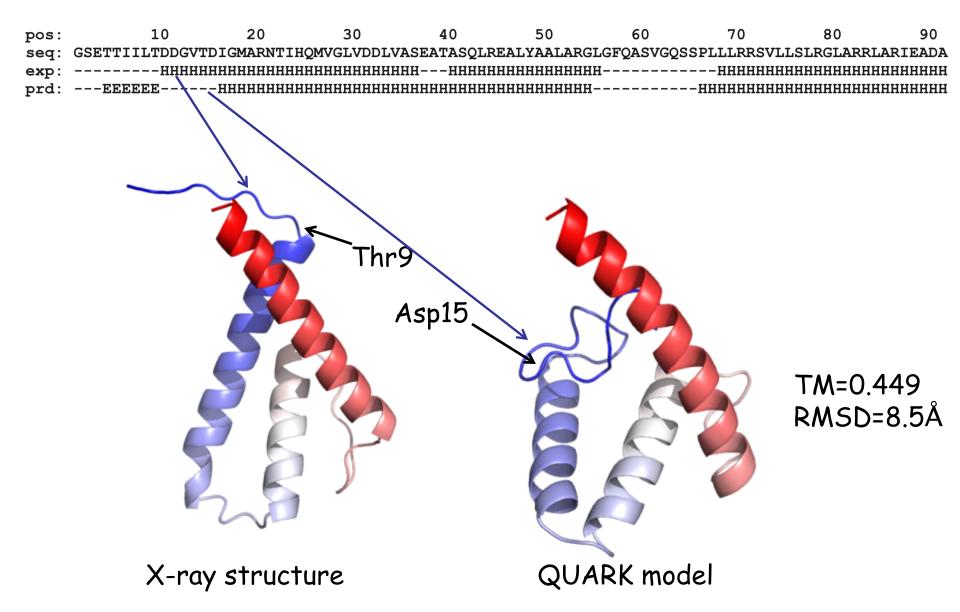
5,527 non-redundant proteins

## 1.4. State of the art: PSSpred Output of PSSpred

Winner takes all, with conf= $10*[P(S_1)-P(S_2)]$ 1 N C 0.015 0.019 0.972 0.010 0.363 0.704 3 V E 0.012 0.705 0.354 4 R E 0.011 0.814 0.250 0.006 1.014 0.063 0.003 1.034 0.043 6 V E 0.007 0.995 0.076 0.013 0.261 0.738 0.009 0.167 0.825 9 G C 0.006 0.964 0.113 10 R E 11 R E 0.006 1.020 0.050 12 V E 0.010 0.998 0.066 13 G E 0.007 0.965 0.103 0.007 0.851 14 W E 0.193 15 V E 0.015 0.754 0.359 Possibility of Possibility of Possibility of coil alpha beta

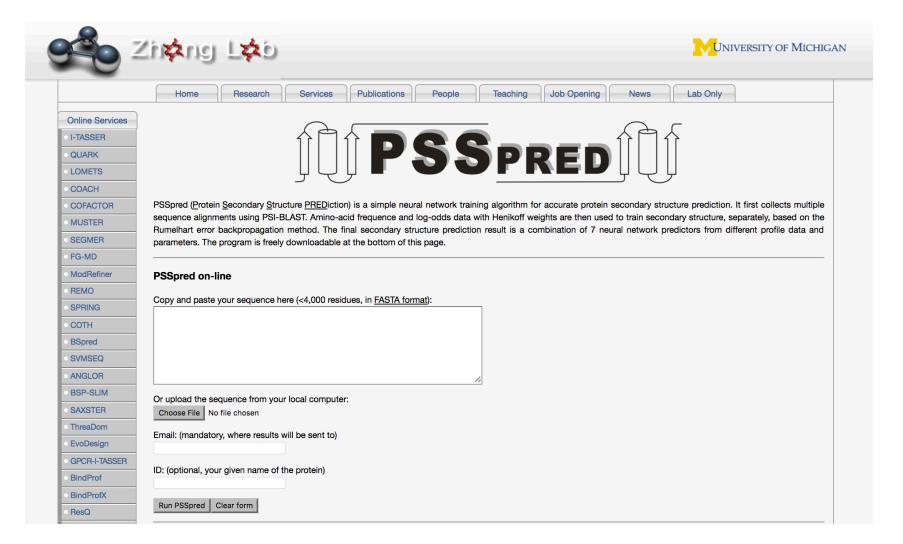
**Result:** Average accuracy =84% on 600 non-redundant proteins, which represents the start of the art of secondary structure prediction. This is close to the experimental uncertainty of hydrogen-bond and SS definition: ~90%.

## Impact of SS prediction on 3D structure prediction (Target T0820-D1 in CASP11)



### The on-line server and standalone program of PSSpred is available at

### http://zhanglab.ccmb.med.umich.edu/PSSpred/



We will discuss it further in Practical Section

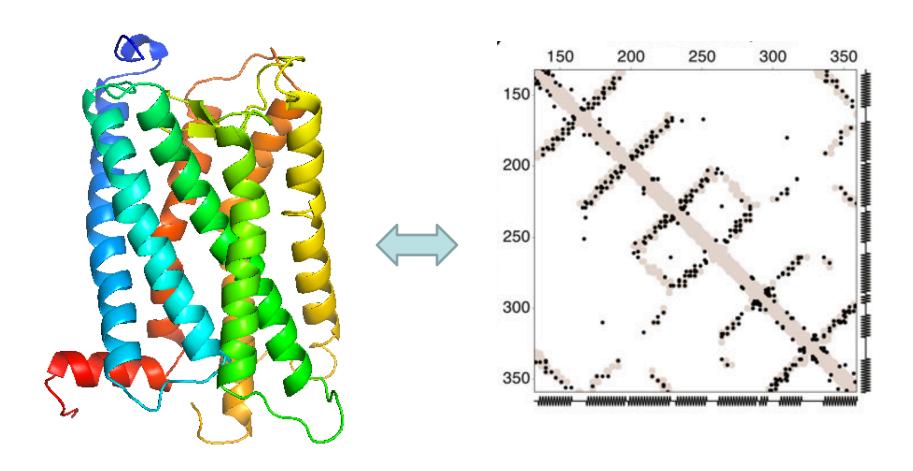
### Conclusions

- 1. Mechine-learning approach could generate the best SS prediction, significantly better than statistical or physical approaches
- 2. Accuracy of NN-based secondary prediction is approaching to its limit of experimental uncertainty. Accordingly, CASP stopped SS prediction competition in CASP5 (2003)
- 3. This study shows that combining multiple predictor algorithms can still give (statistically significant) improvement over individual predictors

### <u>Case Studies of Machine-Learning in</u> <u>Structure Biology</u>

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- 2. Protein Contact Prediction
- 3. Disease-Associated Mutation Prediction

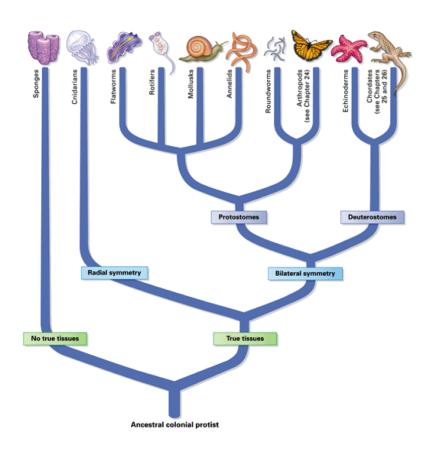
### 2.1 Protein contact-map dictates the 3D fold



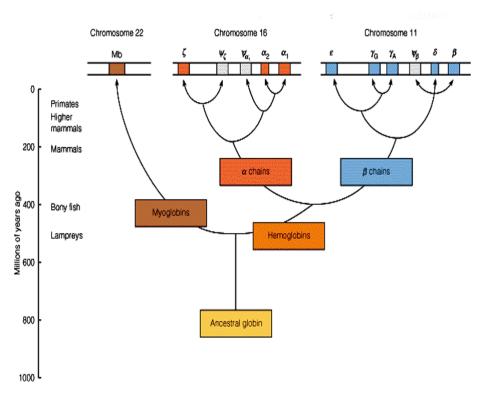
3D structure

2D contact-map

### 2.2. Deriving contact-map from co-evolution coupling

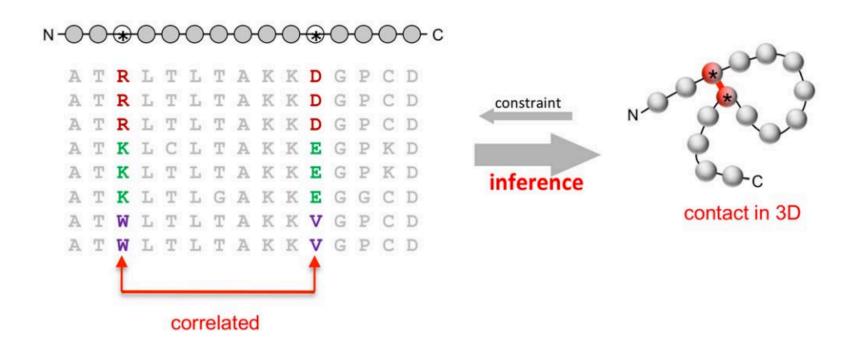


### Globin evolution and expression



### 2.2. Deriving contact-map from co-evolution coupling

Assumption: Spatially contacted residues usually mutate cooperatively



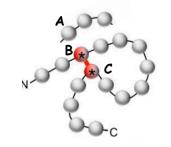
Contacts can be derived by mutual information of  $A_i$ ,  $A_j$ :

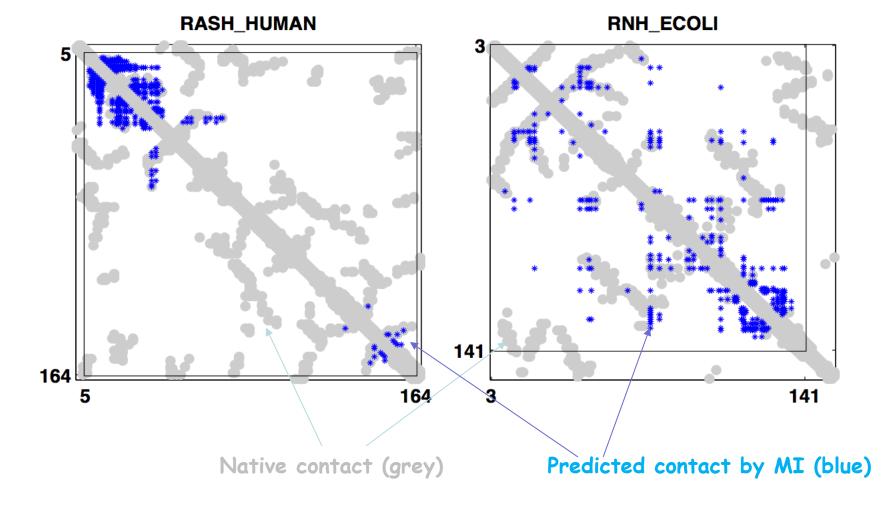
$$MI_{ij} = \sum_{A_i, A_j = 1}^{q} f_{ij}(A_i, A_j) \ln \left( \frac{f_{ij}(A_i, A_j)}{f_i(A_i)f_j(A_j)} \right)$$

### Problem of MI-based co-evolution contact prediction

### **Transitivity issues:**

If  $A \leftarrow B$  and  $B \leftarrow C$ ,  $A \leftarrow C$ ; but Residues A and C are not in contact





### 2.2. Deriving contact-map from Coevolution coupling

Maximum entropy model (Evfold by Marks et al):

Unknown multivariate distribution

**Define:** 
$$P_{ij}(A_i, A_j) \equiv \sum_{\{A_k = 1, ..., q\} k \neq i, j} P(A_1, ..., A_L) = f_{ij}(A_i, A_j)$$

Request: Maximizing entropy of  $P(A_1,...,A_L)$  consistent with data  $f_{ij}(A_i,A_j)$ 

**We have:** 
$$P(A_1,...,A_L) = \frac{1}{Z} \exp \left\{ \sum_{1 \le i \le j \le L} e_{ij} (A_i,A_j) + \sum_{1 \le i \le L} h_i(A_i) \right\}$$

Calculating  $e_{ij}(A_i,A_j)$ :

Direct coupling

$$C_{ij}\left(A_{i},A_{j}\right)=f_{ij}\left(A_{i},A_{j}\right)-f_{i}(A_{i})f_{j}\left(A_{j}\right)$$
 
$$20*20*L*L-dimension$$
 
$$e_{ij}\left(A_{i},A_{j}\right)=-\left(C^{-1}\right)_{ij}\left(A_{i},A_{j}\right)$$

Contacts are predicted from  $DI_{ij}$ :

$$P_{ij}^{Dir}(A_i, A_j) = \frac{1}{Z} \exp\left\{e_{ij}(A_i, A_j) + \tilde{h}_i(A_i) + \tilde{h}_j(A_j)\right\}$$

$$DI_{ij} = \sum_{A_i, A_j = 1}^{q} P_{ij}^{Dir}(A_i, A_j) \ln\left(\frac{P_{ij}^{Dir}(A_i, A_j)}{f_i(A_i)f_j(A_j)}\right)$$

### 2.2. Deriving contact-map from Coevolution coupling

Sparse inverse covariance estimation (PSICOV by Jones):

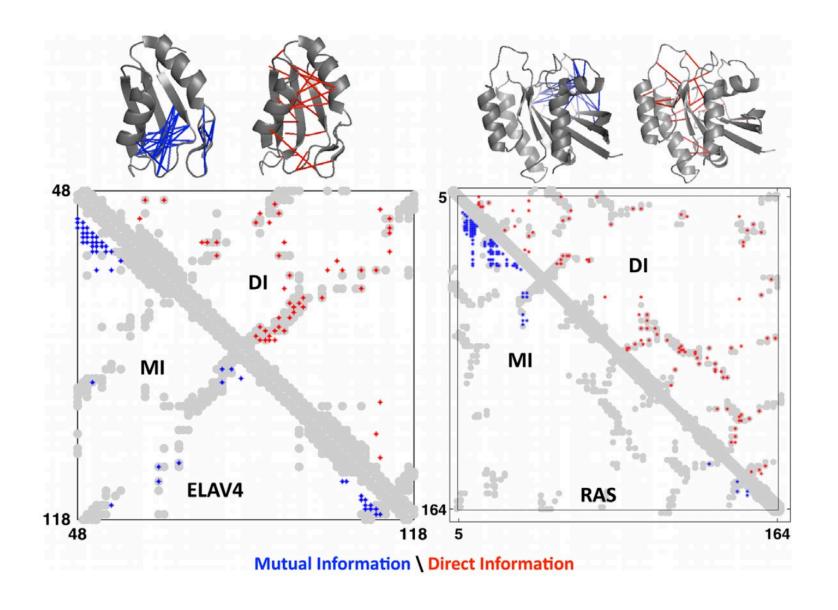
$$PC_{ij} = S_{ij}^{\text{contact}} - \frac{\overline{S}_{(i-)}^{\text{contact}} \overline{S}_{(-j)}^{\text{contact}}}{\overline{S}^{\text{contact}}}$$

$$S_{ij}^{\text{contact}} = \sum_{ab} |\Theta_{ij}^{ab}|$$

$$S_{ij}^{ab} = E(x_i^a x_j^b) - E(x_i^a) E(x_j^b) = f(A_i B_j) - f(A_i) f(B_j)$$

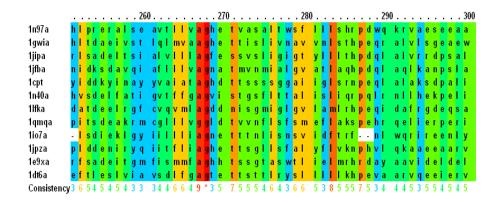
$$\Theta_{ij}^{ab} = (S^{-1})_{ij}(a, b)$$

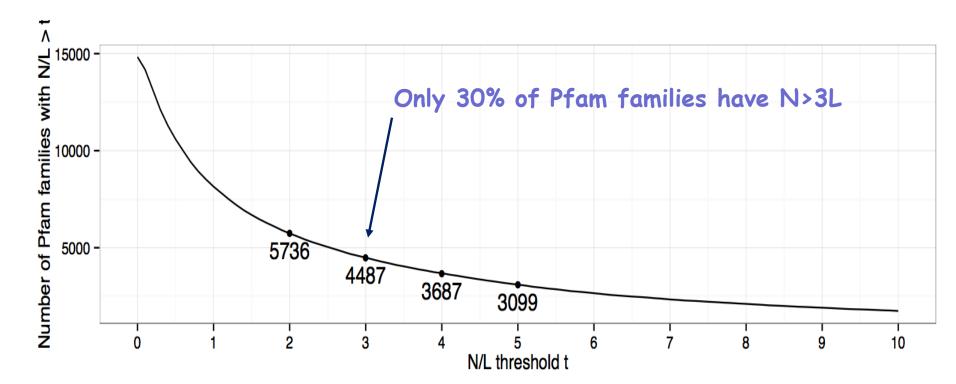
### Direct coupling works better than MI due to noise removal



### Problem of co-evolution contact prediction

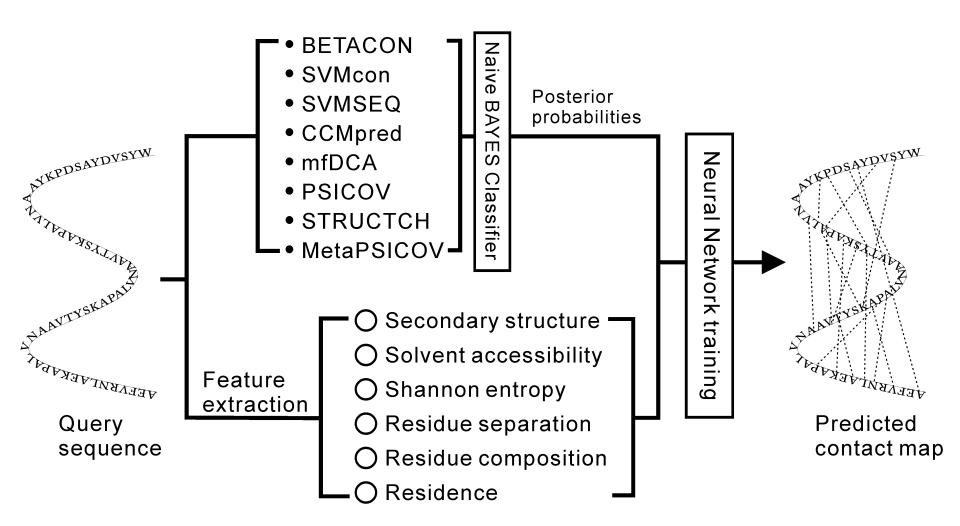
Coevolution based contact predictor works well only when a sufficient number of homologous sequence can be detected, i.e. N>~3\*L





Family statistics for 14831 Pfam families

## 2.3 NeBcon: A new machine-learning approach to contact prediction (by combining neural-network training and naïve Bayes classifier)



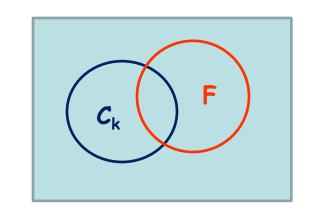
He, Mortuza, Wang, Shen, Zhang, Bioinformatics, 2017

### What is naïve Bayes classifier?

#### Given:

 $F=(f_1, f_2, ..., f_n)$ : n specific features

Ck: k'th possible outcome



### Naïve Bayes classifier theorem:

$$P(C_k|f_1, f_2, \dots, f_n) = P(C_k|F) = \frac{P(C_k)P(F|C_k)}{P(F)}$$

$$P(C_k|F) \propto P(C_k)P(F|C_k) = P(C_k)P(f_1|f_2,\cdots,f_n,C_k)P(f_2|f_3,\cdots,f_n|C_k)\cdots P(f_n|C_k)$$

Under naïve assumption (ie, all features are independent):  $P(f_i|f_{i+1},\cdots,f_n,C_k)=P(f_i|C_k)$ 

$$P(C_k|F) \propto P(C_k) \prod_{i=1}^n P(f_i|C_k)$$
$$P(C_k|F) = \frac{P(C_k) \prod_{i=1}^n P(f_i|C_k)}{P(F)}$$

### An example of application of naïve Bayes classifier

#### Training data:

Sex	height (feet)	weight (lbs)	foot size(inches)
male	6	180	12
male	5.92 (5'11")	190	11
male	5.58 (5'7")	170	12
male	5.92 (5'11")	165	10
female	5	100	6
female	5.5 (5'6")	150	8
female	5.42 (5'5")	130	7
female	5.75 (5'9")	150	9

#### Target sample:

height (feet)	weight (lbs)	foot size(inches)
6	130	8

Question: is this target a male or female?

#### Solution:

Naïve approach: by vote and consensus

Height: male

· Weight: female

· Foot size: female

Conclusion: female

### An example of application of naïve Bayes classifier

#### Training data:

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#### Target sample:

height (feet)	weight (lbs)	foot size(inches)
6	130	8
<b>†</b>	<b>†</b>	<u> </u>
$f_1$	f <sub>2</sub>	f <sub>3</sub>

#### Question: is this target a male or female?

#### Solution:

$$P(C_k|F) = \frac{P(C_k) \prod_{i=1}^n P(f_i|C_k)}{P(F)}$$

$$posterior (male) = \frac{P(\text{male}) \, p(\text{height} \mid \text{male}) \, p(\text{weight} \mid \text{male}) \, p(\text{foot size} \mid \text{male})}{evidence}$$

$$P(\text{male}) = 0.5$$

$$p(\text{height} \mid \text{male}) = \frac{1}{\sqrt{2\pi\sigma^2}} \exp\left(\frac{-(6-\mu)^2}{2\sigma^2}\right) \approx 1.5789$$

$$p(\text{weight} \mid \text{male}) = 5.9881 \cdot 10^{-6}$$

$$p(\text{foot size} \mid \text{male}) = 1.3112 \cdot 10^{-3}$$

$$posterior numerator (male) = \text{their product} = 6.1984 \cdot 10^{-9}$$

$$posterior (\text{female}) = \frac{P(\text{female}) \, p(\text{height} \mid \text{female}) \, p(\text{weight} \mid \text{female}) \, p(\text{foot size} \mid \text{female})}{evidence}$$

$$P(\text{female}) = 0.5$$

$$p(\text{height} \mid \text{female}) = 2.2346 \cdot 10^{-1}$$

$$p(\text{weight} \mid \text{female}) = 1.6789 \cdot 10^{-2}$$

$$p(\text{foot size} \mid \text{female}) = 2.8669 \cdot 10^{-1}$$
The major advantage of NBC over consensus is that the NBC combinate considers specific distribution of individual features.

posterior numerator (female) = their product =  $5.3778 \cdot 10^{-4}$ 

The major advantage of NBC over consensus is that the NBC combination considers specific distribution of individual features.

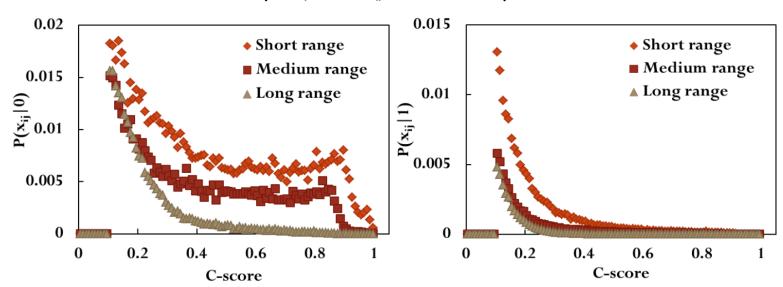
### 2.3 NeBcon: Combining neural-network training and naïve Bayes classifier for protein contact map prediction

Feature type-I: posterior probability of meta-predictors (121 features):

$$P(C|X_{ij}) = \frac{P(C) \prod_{m=1}^{N} P(X_{ij}^{m}|C)}{P(X_{ij})} = \frac{P(C) \prod_{m=1}^{N} P(X_{ij}^{m}|C)}{P(0) \prod_{m=1}^{N} P(X_{ij}^{m}|0) + P(1) \prod_{m=1}^{N} P(X_{ij}^{m}|1)}$$

### Conditional probabilities:

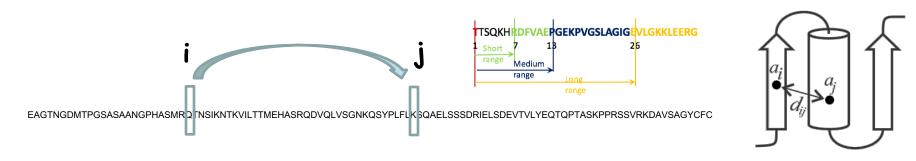
One of the examples (SVMSEQ) for 8 contact predictors:



SVMSEQ: Wu & Zhang, Bioinformatics, 2008

# 2.3 NeBcon: Combining neural-network training and naïve Bayes classifier for protein contact map prediction

Feature type-II: inherent physicochemical feature collection (596 features):



- 1.  $X_i=0,1$  when i'th residue within sequence (22=11×2 features)
- 2. Secondary structure by PSSpred (66=11×2×3 features)
- 3. Solvent accessibility of target residues (22=11×2 features)
- 4. Shannon entropy of i'column in PsiBlast MSA ( $22=11\times2$  features):

$$x_i = \sum_{k=1}^{21} p_k^i ln p_k^i$$

- 5. Sequence separation (2 features): x=|i-j|
- 6. Sequence profile (462=11x2x21 features)

#### 6. Sequence profile (462=11x2x21 features):

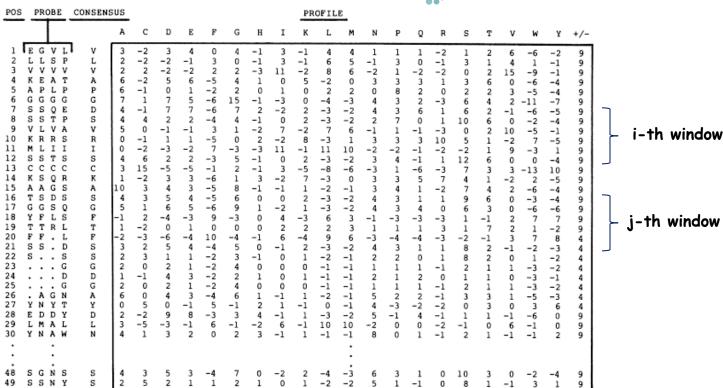
Query sequence: EAGTNGDMTPGSASAANGPHASMRQTNSIKNTKVILTTMEHAS

RQDVQLVSGNKQSYPLFLKSQAELSSSDRIELSDEVTVLYEQTQ

PTASKPPRSSVRKDAVSAGYCFC



#### Sequence profile:

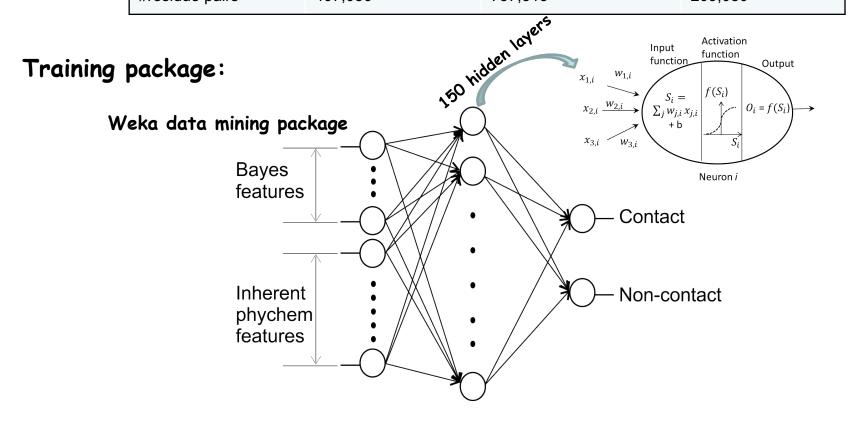


### Neural Network Training

#### Training set:

517 non-homologous proteins containing:

	Short-range  i-j <7	Medium range 6< i-j <24	Long-range 23< i-j
#true contacts	20,636	26,798	87,200
#residue pairs	407,036	757,315	209,080



Hall et al. The WEKA data mining software: an update. SIGKDD Explor. Newsl., 11, 10 (2009).

### Test Results

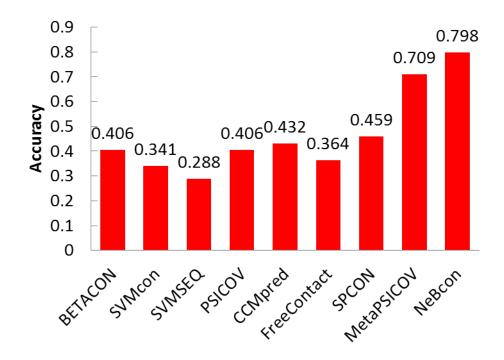
#### Test protein set:

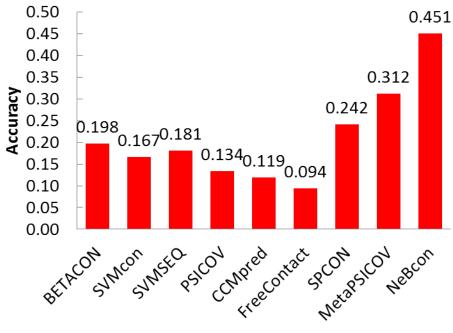
98 proteins containing 3850, 5849, 13792 short-, medium- and long-range contacts

Accuracy of the prediction:  $Acc = N_{corr}/N_{T}$ 

- $\bullet N_{corr} = \#$  of correctly predicted contacts in the contact map
- $\bullet N_T = \#$  of predicted contacts in the contact map

#### Results:





50 easy targets
Top L/5 long range

48 hard targets
Top L/5 long range

## Test Results

#### Combine all target together:

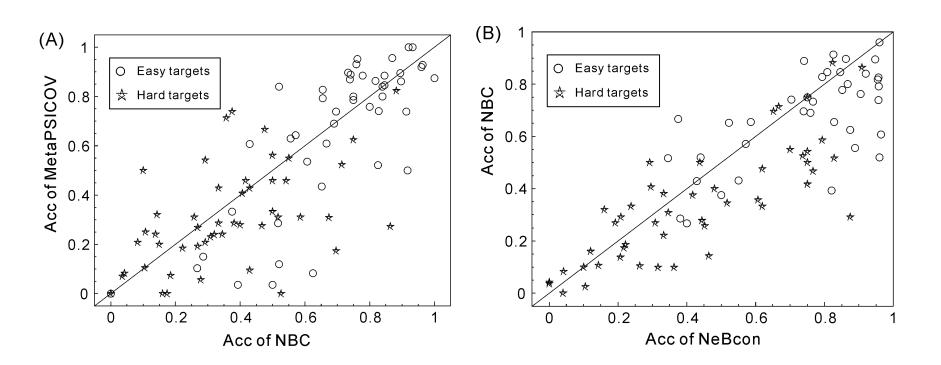


**Table 2.** Average accuracy of top L/5 contact predictions by different methods on 98 test proteins

Methods	Short (6–11)	Medium (12–24)	Long (>24)
BETACON	0.540 (1*10 <sup>-9</sup> )	0.430 (3*10 <sup>-10</sup> )	$0.310 (2*10^{-12})$
SVMSEQ	$0.475 (2*10^{-12})$	$0.393 (2*10^{-12})$	$0.255 (2*10^{-12})$
SVMcon	$0.564 (4*10^{-9})$	$0.455 (1*10^{-8})$	$0.236 (2*10^{-12})$
PSICOV	$0.204 (2*10^{-12})$	$0.246 (2*10^{-12})$	$0.262 (2*10^{-12})$
CCMpred	$0.206 (2*10^{-12})$	$0.238 (2*10^{-12})$	$0.278 (2*10^{-12})$
FreeContact	0.234 (2*10-12)	$0.278 (2*10^{-12})$	$0.232 (2*10^{-12})$
STRUCTCH	$0.605 (3*10^{-4})$	$0.487 (4*10^{-5})$	$0.353 (2*10^{-12})$
MetaPSICOV	$0.576 (5*10^{-6})$	$0.572 (5*10^{-1})$	$0.515 (2*10^{-7})$
NeBcon	0.651	0.574	0.628

NeBcon significantly outperforms all individual contact predictors

## Comparison of NeBcon to the best predictor



- 1. Bayes combination contributes to overall performance
- 2. NN training increases accuracy for hard targets that have low number of homologous sequences

## Testing results on CASP targets

### Contact prediction on the free-modeling (FM) targets in CASP

CASP10 (20 FM tar	gets)	CASP11 (33 FM targets)				
Methods	Accuracy (p-value)	Methods	Accuracy (p-value)			
NeBcon	0.4659	NeBcon	0.3763			
Multicon	$0.4058 (2.5 \times 10^{-1})$	MetaPSICOV	$0.3632 (3.9 \times 10^{-1})$			
Distill_roll	$0.2804 (7.5 \times 10^{-3})$	Pcons-net	$0.2482 (1.0 \times 10^{-3})$			
Distill	$0.2448 (3.5 \times 10^{-3})$	Shen-group	$0.2330 (4.5 \times 10^{-3})$			
Multicon-Const	$0.2252 (4.0 \times 10^{-4})$	UCI-IGB-CMpro	0.2199 (4.8×10 <sup>-3</sup> )			
IGBteam	$0.2038 (1.3 \times 10^{-4})$	RBO_Aleph	$0.1990 (2.9 \times 10^{-3})$			
SAM-T08-server	$0.1924 (1.8 \times 10^{-4})$	LEE	$0.1988 (1.1 \times 10^{-3})$			
MetaPSICOV	$0.1721 (4.5 \times 10^{-5})$	Multicom-Clust	0.1831 (4.1×10 <sup>-4</sup> )			
RaptorX-Roll	$0.1573 (6.2 \times 10^{-5})$	RaptorX-Contact	0.1605 (7.2×10 <sup>-5</sup> )			
Multicon-Novel	$0.1514 (7.6 \times 10^{-6})$	Multicon-Const	0.1559 (4.7×10 <sup>-5</sup> )			
ZHOU-SPARKS-X	$0.0864 (3.0 \times 10^{-6})$	Distill	0.0782 (2.7×10 <sup>-6</sup> )			

Evenness of contact-map distributed

Shannon entropy of predicted contact-map

**Methods** 

**BETACON** 

**SVMSEQ** 

**SVMcon** 

**PSICOV** 

**CCMpred** 

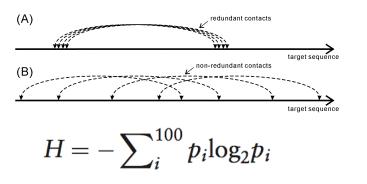
FreeContact

**STRUCTCH** 

**MetaPSICOV** 

NeBcon

Native



2.705 (2.2\*10<sup>-9</sup>)

 $2.680 (5.6*10^{-15})$ 

 $2.589 (5.7*10^{-16})$ 

 $2.676 (2.6*10^{-2})$ 

 $3.377 (1.3*10^{-7})$ 

 $3.245 (2.0*10^{-4})$ 

 $2.723 (1.5*10^{-9})$ 

 $2.958 (3.8*10^{-1})$ 

 $2.750 (5.6*10^{-10})$ 

2.973

**Short** 

30	30 60 90 Residue Order	*** *** 12
Long	All	
2.656 (8.4*10 <sup>-16</sup> )	3.912 (6.9*10 <sup>-25</sup> )	
3.540 (4.9*10 <sup>-7</sup> )	4.146 (5.6*10 <sup>-13</sup> )	
3.289 (1.5*10 <sup>-16</sup> )	3.962 (1.2*10 <sup>-24</sup> )	
3.505 (6.2*10 <sup>-2</sup> )	3.959 (1.23*10 <sup>-2</sup> )	
4.415 (6.9*10 <sup>-9</sup> )	5.016 (1.1*10 <sup>-6</sup> )	
4.478 (4.5*10 <sup>-10</sup> )	4.977 (5.0*10 <sup>-6</sup> )	
3.477 (2.6*10 <sup>-8</sup> )	4.072 (7.7*10 <sup>-17</sup> )	
3.552 (4.0*10 <sup>-5</sup> )	4.217 (9.7*10 <sup>-6</sup> )	
		i

 $4.273 (3.3*10^{-9})$ 

4.473

sidue Order

 $3.665 (6.5*10^{-5})$ 

3.815

Most predictors create contact map with similar evenness as native

2.823

**Medium** 

1.953 (5.2\*10<sup>-18</sup>)

 $2.523 (2.0*10^{-8})$ 

 $2.402 (5.2*10^{-13})$ 

 $2.726 (2.6*10^{-1})$ 

 $3.472 (8.3*10^{-13})$ 

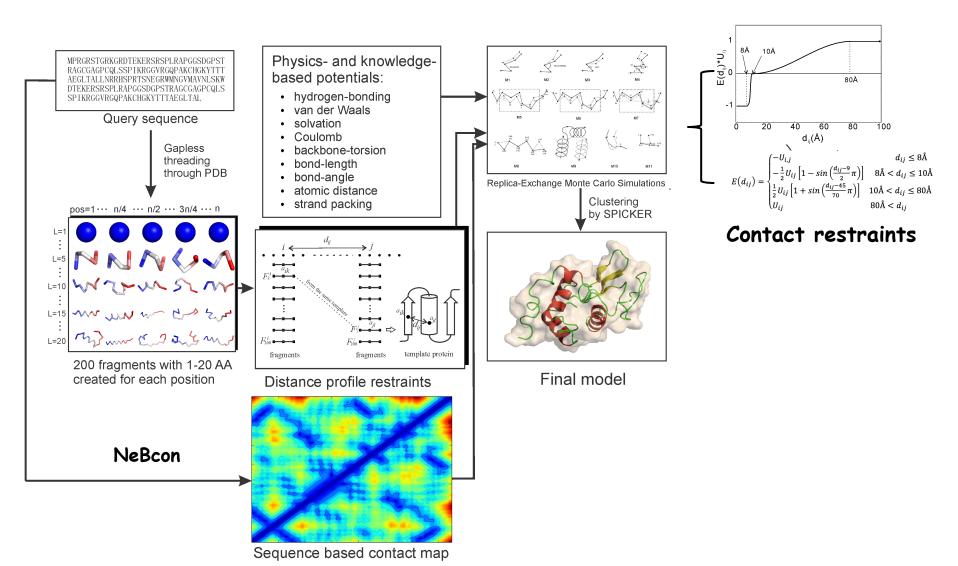
 $3.426 (2.1*10^{-11})$ 

 $2.647 (3.0*10^{-4})$ 

 $2.709 (1.4*10^{-2})$ 

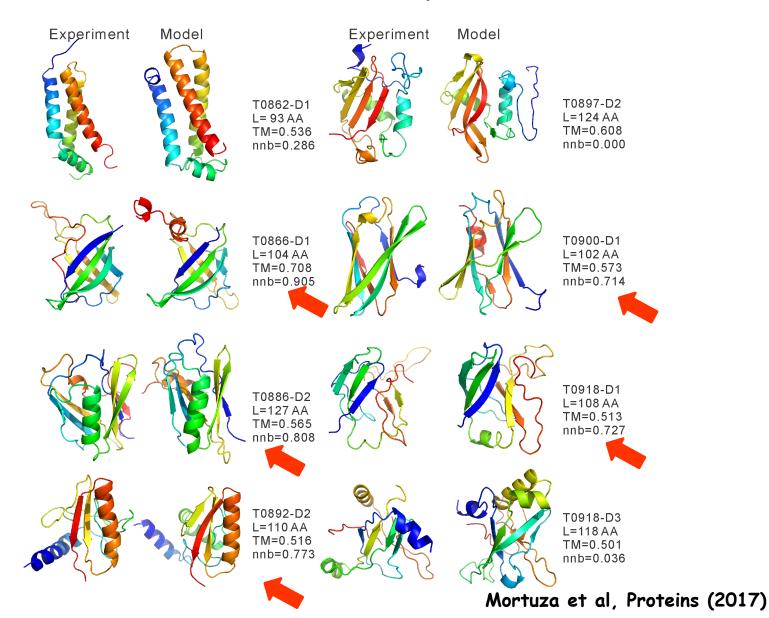
 $2.570 (7.8*10^{-10})$ 

# C-QUARK: Using contact-map prediction to guide ab initio protein structure folding in CASP12

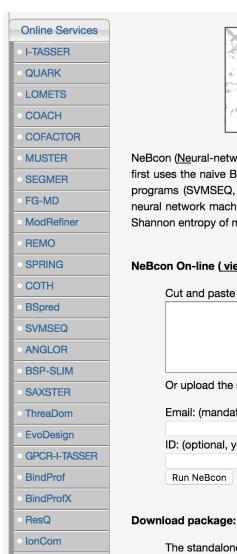


Xu, Zhang. Proteins (2012)

# 5 of 8 successful *ab initio* folding cases in CASP12 are due to contact prediction



#### NeBcon is freely available at <a href="http://zhanglab.ccmb.med.umich.edu/NeBcon/">http://zhanglab.ccmb.med.umich.edu/NeBcon/</a>





NeBcon (Neural-network and Bayes-classifier based contact prediction) is a hierarchical algorithm for sequence-based protein contact map prediction. It first uses the naive Bayes classifier theorem to calculate the posterior probability of eight machine-learning and co-evoluation based contact prodiction programs (SVMSEQ, BETACON, SVMcon, PSICOV, CCMpred, FreeContact, MetaPSICOV, and STRUCTCH). Final contact maps are then created by neural network machine that trains the posterior probability scores with intrinsic structural features from secondary structure, solvent accessibility, and Shannon entropy of multiple sequence alignments.

The standalone NeBcon package can be downloaded from NeBconpackage.tar.gz. In order to install and run the package, follow the instrunctions

#### NeBcon On-line (view an example of NeBcon output)

Cut and paste your sequence (in FASTA format) below:
Or upload the sequence from your local computer: Choose File No file chosen
Email: (mandatory, where results will be sent to)
ID: (optional, your given name of the protein)
Run NeBcon Clear form

We will discuss on its application in Pratical Section

### **Conclusions**

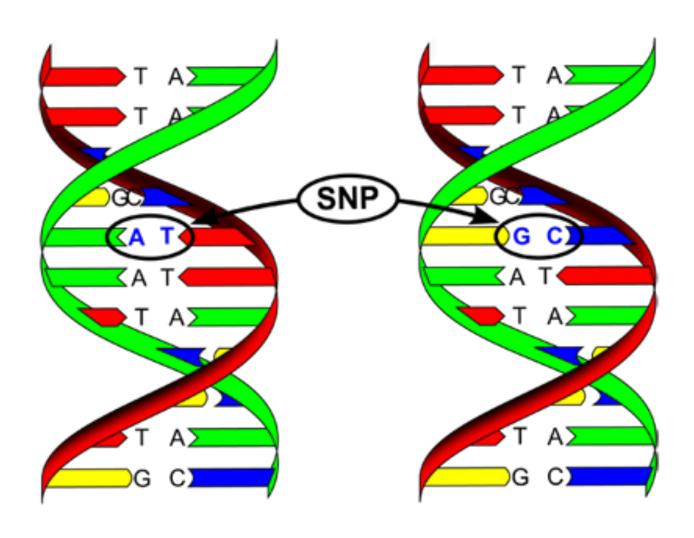
- 1. Contact prediction can improve accuracy of ab initio protein structure for targets without templates. This is particularly true given (a) sequence library increases; (b) new methods for removing translation correlation
- 2. Naïve Bayes classifier helps combining multiple contact predictors
- 3. NN training on inherent protein features improves contact prediction for hard targets

## <u>Case Studies of Machine-Learning in</u> <u>Structure Biology</u>

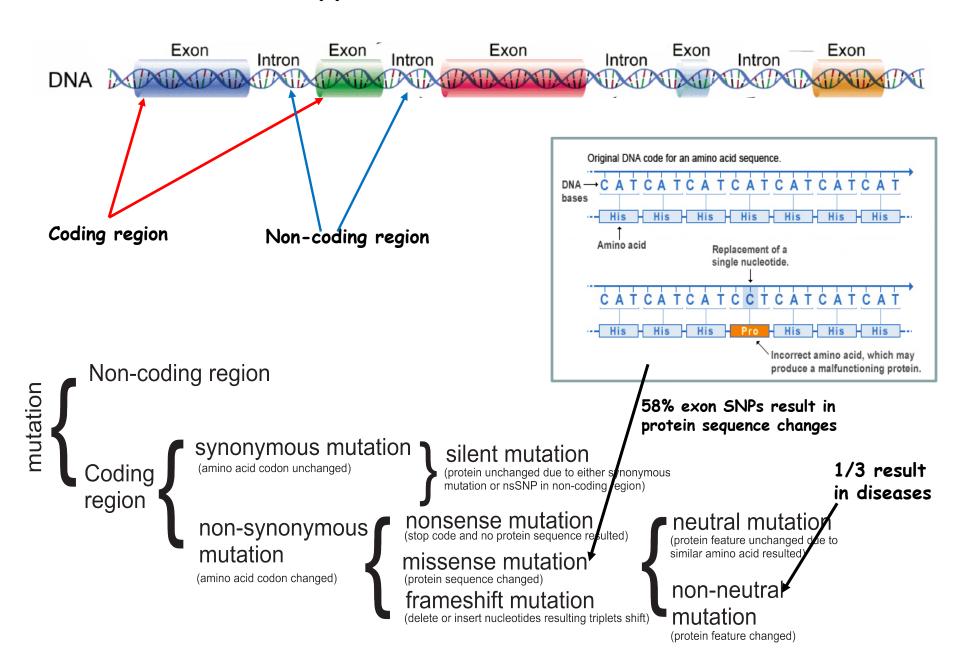
- 1. Protein Secondary Structure Prediction
- 2. Protein Contact Prediction
- 3. Disease-Associated Mutation Prediction

## Single-nucleotide polymorphism (SNP)

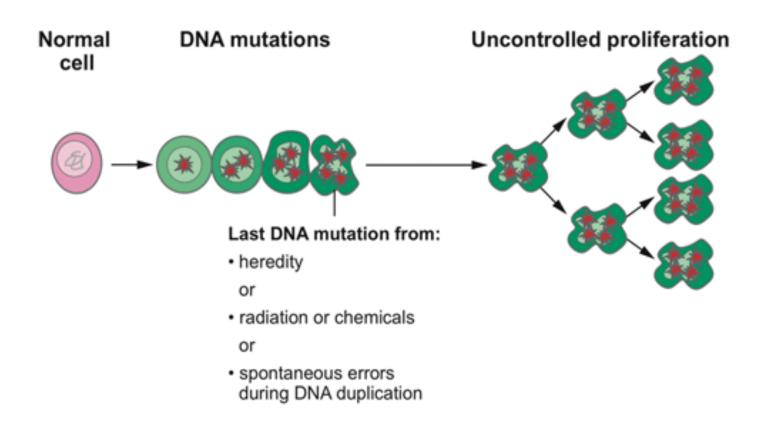
Genome evolution is mainly driven by SNP mutations



### Types of SNP mutations



## Cancer Arises From DNA Mutations in Cells



## Genetic Diseases

More than 6,000 diseases are due to SNP mutations

- · cystic fibrosis (囊胞性纤维症)
- · sickle cell anemia (镰状细胞性贫血)
- · Marfan syndrome (马方综合征)
- · Huntington's disease (亨廷顿氏舞蹈病)
- · Hemochromatosis (血色沉着病) 👡







Some serious diseases due to mutation on multiple genes:

- · heart disease (心脏病)
- · high blood pressure (高血压)
- · Alzheimer's disease (早老性痴呆)
- · Arthritis (关节炎) ←
- · Diabetes (糖尿病)
- · Obesity (肥胖) 🖊
- · Cancer (癌症)





How to predict what mutations could cause diseases and what could not?

PreDAM: predicting disease-associated mutations based on machine learning



http://zhanglab.ccmb.med.umich.edu/PreDAM/

### Group-I: Physicochemical properties (Pharmacophore)

associated mutations

mutations

H-bond acceptor	
-----------------	--

Feature	ture No. Feature							Description	H-bond donor	
Class			DM <sup>a</sup>	NM <sup>b</sup>	cutoff	value	W Test) <sup>c</sup>			
Physico-	Phar	macophore	for the wild-	type residues					Anionic	000
chemical	1	$HP_{w}$	2.909	2.23	2	0.14	6.17E-53	Hydrophobic		
properties	2	$noHP_w$	4.479	3.98	6	0.11	2.11E-25	Non hydrophobic	WO.2007 VIOLET CO. 100 VIOLET CO. 10	~~~
	3	$AR_{w}$	1.289	0.99	2	0.11	1.13E-29	Aromatic rings	Cationic	and the
	4	$noAR_w$	6.09	5.23	6	0.15	1.60E-63	Non aromatic rings		
	5	$PC_{w}$	0.869	0.80	3	0.03	3.30E-3	Positive charge	Lludrophobio	
	6	$NC_w$	0.72	0.76	4	0.05	1.93E-2	Negative charge	Hydrophobic	
	7	$noC_w$	5.79	4.66	5	0.19	1.12E-89	Neutral charge		a #
	8	$\mathrm{BP}_{\mathrm{w}}$	1.67	1.27	2	0.08	6.73E-9	Both wild-type and neighbor AA are polar	Aromatic	0
	9	$OP_w$	2.83	2.29	3	0.11	2.33E-39	Either of wild-type and neighbor AA is polar		
	10	$NP_{w}$	1.87	1.65	5	0.08	2.69E-3	Both wild-type and neighbor are nonpolar t		8
	11	$AC_w$	9.12	8.05	11	0.11	1.16E-18	The count of reside being the hydrogen accep-	O1	0
	12	$\mathrm{DO}_{\mathrm{w}}$	5.59	4.86	8	0.13	1.73E-25	The count of reside being the hydrogen donor		
	Pharmacophore for the mutant residues									
	13	HP <sub>m</sub>	3.04	2.30	3	0.16	8.54E-62	Hydrophobic		
	14	$noHP_m$	4.52	3.79	5	0.16	4.78E-56	Non hydrophobic		
	15	$AR_m$	1.33	1.03	1	0.10	2.96E-29	Aromatic rings		
	16	$noAR_m$	6.23	5.06	7	0.19	3.5E-104	Non aromatic rings		
	17	$PC_m$	0.83	0.68	2	0.07	2.63E-11	Positive charge		
	18	$NC_m$	0.69	0.69	4	0.03	1.88E-1	Negative charge		
	19	$noC_m$	6.041	4.72	5	0.19	1.7E-105	Neutral charge		
	20	$BP_m$	1.38	1.22	4	0.05	1.1E-1	Both wild-type and neighbor AA are polar		0=0
	21	$OP_m$	3.22	2.37	5	0.17	8.09E-73	Either of wild-type and neighbor AA is polar		1
	22	$NP_m$	1.95	1.50	5	0.12	1.75E-14	Both wild-type and neighbor are nonpolar t		
	23	$AC_m$	9.31	7.84	15	0.14	9.92E-30	The count of reside being the hydrogen accept	or	
	24	$\mathrm{DO}_{\mathrm{m}}$	5.69	4.79	7	0.14	7.39E-32	The count of reside being the hydrogen donor		
r			1	<u> </u>			1			
		1	Disease	Neutral			P-value in			

Mann-

White test

#### Group-I: Physicochemical properties (contact environments)

Feature	No.	Feature	M	ean	M	CC	p-value (M-	Description
Class			DM <sup>a</sup>	NM <sup>b</sup>	cutoff	value	W Test) <sup>c</sup>	
Physico-								
chemical	Muto	ation-induced	environmer	ıtal pharmac	ophore cha			
properties	25	$\cos_{\mathrm{WM}}$	0.74	0.80	0.89	0.17	5E-66	The cosin for the pharmacophores of wild-type and mutant residues
	26	$rms_{WM}$	0.47	0.39	0.41	0.17	1.14E-61	The RMSD for the pharmacophores of wild-type and mutant residues
	27	$\cos N_{WM}$	0.94	0.95	0.97	0.06	1.25E-07	The cosin for the neighbor pharmacophores of wild-type and mutant residues
	28	$rmsN_{WM} \\$	2.09	1.58	1.68	0.19	6.91E-98	The RMSD for the neighbor pharmacophores of wild-type and mutant residues
	29	cosNS <sub>WM</sub>	0.92	0.93	1.00	0.08	5.48E-11	The cosin for the neighbor pharmacophores of wild-type and mutant residues related with single residue
	30	$rmsNS_{WM} \\$	1.77	1.30	1.84	0.21	1.2E-110	The RMSD for the neighbor pharmacophores of wild-type and mutant residues related with single residue
	31	cosNP <sub>WM</sub>	0.92	0.93	1.00	0.04	2.08E-1	The cosin for the neighbor pharmacophores of wild-type and mutant residues related with residue paired
	32	$rmsNP_{WM}$	2.87	2.27	4.24	0.12	6.87E-28	The RMSDfor the neighbor pharmacophores of wild-type and mutant residues related with residue paired
	Othe	r physicoche	mical proper	rties				
	33	Volw	2.83	2.86	1.90	0.09	4.51E-1	The volume of wild-type residue
	34	Volm	2.91	2.89	3.16	0.09	1.86E-3	The volume of mutant residue
	35	dVol	0.08	0.03	0.65	0.13	2.13E-05	The volume difference
	36	Ww	132.01	131.84	75.07	0.09	0.19646	The weight of wild-type residue
	37	Wm	136.24	133.09	165.19	0.09	8.7E-05	The weight of mutant residue
	38	dW	4.23	1.26	42.08	0.15	4.78E-4	The molecular weight difference

$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{p_{w}(k)} = \{P_{w}^{1}(i), \dots, P_{w}^{L}(i)\}$$

$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{p_{w}(k)} = \{P_{w}^{1}(i), \dots, P_{w}^{L}(i)\}$$

$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{p_{w}(\iota)} = \{P_{w}^{1}(i), \dots, P_{w}^{L}(i)\}$$

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$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{P_{w}(\iota)} = \{P_{w}^{1}(\iota), \dots, P_{w}^{L}(\iota)\}$$

$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{P_{w}(\iota)} = \{P_{w}^{1}(\iota), \dots, P_{w}^{L}(\iota)\}$$

$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{P_{w}(\iota)} = \frac{\overrightarrow{P_{w}(\iota)} \cdot \overrightarrow{P_{w}(\iota)}}{\|\overrightarrow{P_{w}(\iota)}\| \cdot \|\overrightarrow{P_{w}(\iota)}\|}$$

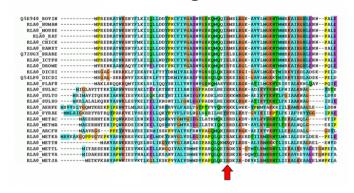
$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{P_{w}(\iota)} = \{P_{w}^{1}(\iota), \dots, P_{w}^{L}(\iota)\}$$

$$\overrightarrow{P_{w}(\iota)} = \sum_{k=1}^{n_{con}^{w}(\iota)} \overrightarrow{P_{w}(\iota)} = \{P_{w}^{1}(\iota), \dots, P_{w}^{L}(\iota)\}$$

#### Group-II: Evolutionary profiles (PSIBlast, LOMETS, Pfam families):

Feature	No.	Feature	Me	ean	M	CC	p-value (M-	Description
Class			<b>DM</b> <sup>a</sup>	$NM^{b}$	cutoff	value	W Test) <sup>c</sup>	
Evoluti	PSI-	BLAST profil	e scores					
onary	39	$PSIC_w$	1.567	0.99	1.33	0.37	0	The PSIC score for wild-type residue
profiles	40	PSIC <sub>m</sub>	-0.42	0.17	-0.11	0.39	0	The PSIC score for mutant residue
	41	dPSIC	-1.99	-0.82	-1.10	0.48	0	The PSIC score difference
	42	$\mathrm{JSD}_{\mathrm{w}}$	0.03	0.03	0.04	0.10	4.57E-1	The JSD score for wild-type residue
	43	$JSD_m$	0.02	0.02	0.03	0.09	2.74E-07	The JSD score for mutant residue
	44	dJSD	-0.00	-0.00	-0.01	0.08	3.19E-06	The JSD score difference
	45	$JSD_i$	0.47	0.33	0.46	0.29	2.4E-200	The JSD score at mutant position i
	LOM	METS profile s	scores					
	46	$tPSIC_w$	0.78	0.46	0.74	0.20	7.5E-118	The PSIC score for wild-type residue
	47	tPSIC <sub>m</sub>	-0.30	-0.08	0.04	0.15	1.2E-58	The PSIC score for mutant residue
	48	dtPSIC	-1.08	-0.54	-0.67	0.25	1.1E-159	The PSIC score difference
	Pfan	n profile score	S					
	49	$Pfam_w$	1.83	2.35	1.49	0.29	1.1E-124	The Pfam score for wild-type residue
	50	Pfam <sub>m</sub>	3.66	3.10	3.12	0.25	2E-119	The Pfam score for mutant residue
	51	dPfam	1.83	0.75	1.12	0.32	2.6E-190	The Pfam score difference

#### Jensen-Shannon divergence

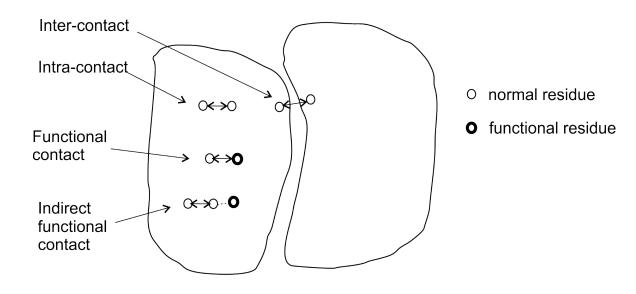


#### Measuring evolutionary divergence

$$\begin{cases} JSD_{ia} = \lambda p_{ia}log \frac{p_{ia}}{c_{ia}} + (1 - \lambda)q_{a}log \frac{q_{a}}{c_{ia}} \\ JSD_{i} = \sum_{a \in AA} JSD_{ia} \end{cases}$$

#### Group-III: Contact environments with functional residues:

Feature	No.	Feature	Mea	n	MCC		p-value (M-	Description
Class			<b>DM</b> <sup>a</sup>	NM <sup>b</sup>	cutoff	value	W Test) <sup>c</sup>	
Contact	Dire	ctly contacted	residues					
environme	52	Intra	13.78	11.25	15	0.23	9.5E-137	The number of intramolecular contacts
nts	53	FunIntra	4.70	3.72	16	0.10	1.35E-15	The number of intramolecular functional contacts
	54	Inter	1.16	0.89	1	0.10	1.96E-15	The number of intermolecular contacts
	55	FunInter	0.30	0.38	5	0.03	3.53E-1	The number of intermolecular functional contacts
	Indi	ectly contacted	d residues					
	56	CIntra	57.96	46.18	66	0.24	5E-132	The number of intramolecular indirectly contacts
	57	CFunIntra	18.67	15.00	1	0.09	7.57E-18	The number of intramolecular functional indirectly contacts
	58	CInter	11.54	8.05	11	0.11	8.59E-18	The number of intermolecular indirectly contacts
	59	CFunInter	3.95	3.23	1	0.10	5.39E-13	The number of intermolecular functional indirectly contacts

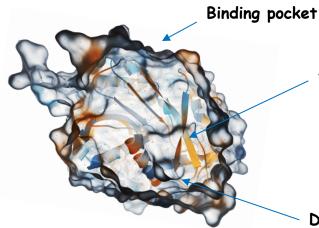


#### Group-IV: Structure prediction based features:

Feature	No.	Feature	M	ean	MO	CC	p-value (M-	Description
Class			<b>DM</b> <sup>a</sup>	NM <sup>b</sup>	cutoff	value	W Test) <sup>c</sup>	
I-	Prote	ein surface re	gions favora	able for intera	ctions			
TASSER	60	CS	0.08	0.04	0.04	0.16	6.46E-56	the ConCavity score for the ligand-binding interaction
structure	61	Depth	6.72	5.50	5.61	0.24	4.2E-137	The average distance of target atoms/residues to its closest
models								molecule of bulk solvent
	The p	hysics-based	energy func	tions				
	62	ED	649.56	589.01	555.57	0.12	8.93E-18	The EvoDesign score
	63	ddG	1.62	0.62	3.00	0.19	4.27E-61	The free-energy changes upon mutation
	64	$\mathrm{VDW}_{\mathrm{w}}$	-343.44	-322.54	-357.52	0.11	1.39E-9	Van der Waals potential of wild-type residue by CISS-RR
	65	$VDW_m$	-331.33	-316.17	-351.06	0.09	5.18E-6	Van der Waals potential of mutant residue by CISS-RR
	66	dVDW	12.11	6.37	1.49	0.20	1.43E-96	Van der Waals potential difference
	67	$RT_{w}$	460.13	424.66	425.50	0.08	1.75E-11	Rotamer preferences of side-chain conformers by CISS-RR.
	68	$CISRR_w$	116.69	102.12	55.05	0.10	2.78E-13	CIS-RR score for wild-type residue
	69	$CISRR_m$	129.06	108.74	67.00	0.13	6.98E-25	CIS-RR score for mutant residue
	70	dCISRR	12.37	6.61	4.52	0.17	1.18E-72	CIS-RR score difference

#### sequence

FTVSNTNNEFVLISDP TGGKSIGLLCFRQED AEAFLAQARLRRREL KTNAKVVPITLDQVYL LKVEGISFRFLPDPI I-TASSER



Atomic interactions

Depth of atoms

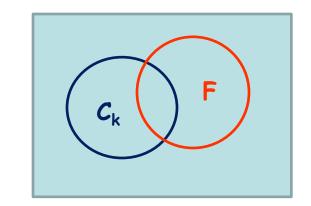
### BANN: A new method for NN training on posterior probability

#### Given:

 $C_{\rm D}$ : disease-associate mutation

 $C_N$ : neutral mutation without causing disease

 $F=(f_1, f_2, ..., f_n)$ : n specific features



#### Naïve Bayes classifier theorem:

$$P(C_k|f_1, f_2, \dots, f_n) = P(C_k|F) = \frac{P(C_k)P(F|C_k)}{P(F)}$$

$$P(C_k|F) \propto P(C_k)P(F|C_k) = P(C_k)P(f_1|f_2,\cdots,f_n,C_k)P(f_2|f_3,\cdots,f_n|C_k)\cdots P(f_n|C_k)$$

Under naïve assumption (ie all features are independent):  $P(f_i|f_{i+1},\cdots,f_n,C_k)=P(f_i|C_k)$ 

$$P(C_k|F) \propto P(C_k) \prod_{i=1}^n P(f_i|C_k)$$

$$\log P(C_k|F) \propto \sum_{i=1}^n \log P(f_i|C_k) + \log P(C_k)$$

### BANN: A new method for NN training on posterior probability

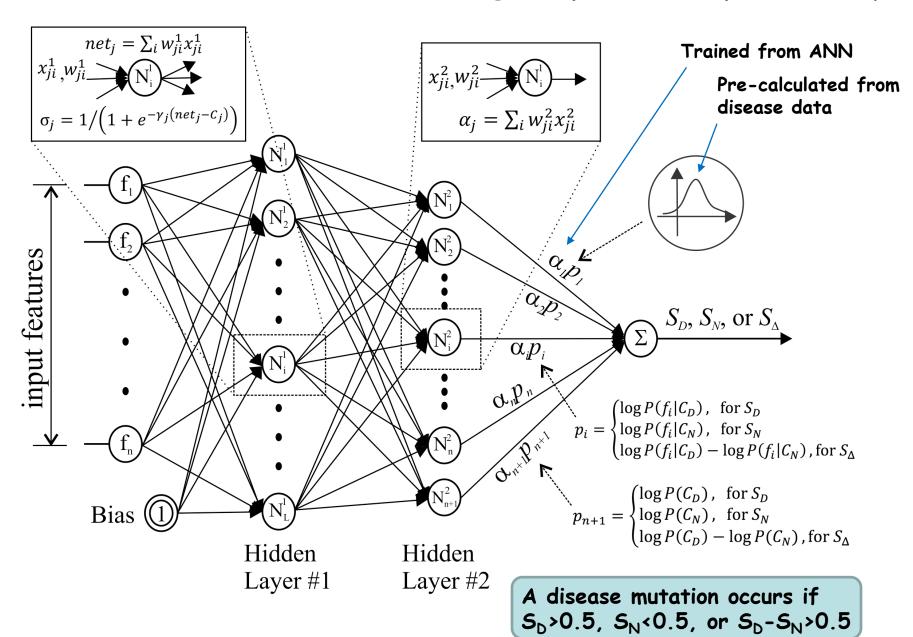
#### General form of Bayes classifier for mutation classes:

$$\log P(C_k|F) = \sum_{i=1}^{n} \alpha_i(F) \log P(f_i|C_k) + \alpha_{n+1}(F) \log P(C_k)$$

$$\begin{cases} S_{D} = \log P(C_{D}|F) = \sum_{i=1}^{n} \alpha_{i}(F) \log P(f_{i}|C_{D}) + \alpha_{n+1}(F) \log P(C_{D}) \\ S_{N} = \log P(C_{N}|F) = \sum_{i=1}^{n} \alpha_{i}(F) \log P(f_{i}|C_{N}) + \alpha_{n+1}(F) \log P(C_{N}) \end{cases}$$

$$S_{\Delta} = S_{D} - S_{N} = \sum_{i=1}^{n} \alpha_{i}(F) [\log P(f_{i}|C_{D}) - \log P(f_{i}|C_{N})] + \alpha_{n+1}(F) [\log P(C_{D}) - \log P(C_{N})].$$

### A new method for NN training on posterior probability



## ANN: back propagation training

#### Error minimization

$$E(\vec{w}, \vec{\gamma}, \vec{C}) \equiv \frac{1}{2N_d} \sum_{d=1}^{N_d} (t_d - o_d)^2 + \mu \sum_{i,j} w_{ji}^2$$

#### Gradient descent training rules

$$\begin{cases} \Delta w_{ji,2} = \eta \delta_{j,2} x_{ji} - 2\eta \mu w_{ji} \\ \Delta w_{ji,1} = \eta \delta_{j,1} x_{ji} - 2\eta \mu w_{ji} \\ \Delta \gamma_{j} = \eta \delta_{j,1} (net_{j} - C_{j}) \\ \Delta C_{j} = \eta \delta_{j,1} \gamma_{j} \end{cases} \begin{cases} \delta_{j,2} = P_{j}(t_{d} - o_{d}) \\ \delta_{j,1} = \gamma_{j} o_{j} (1 - o_{j}) \sum_{k \in Down(j)} \delta_{k,2} w_{kj} \end{cases}$$

learning rate

### Benchmark results

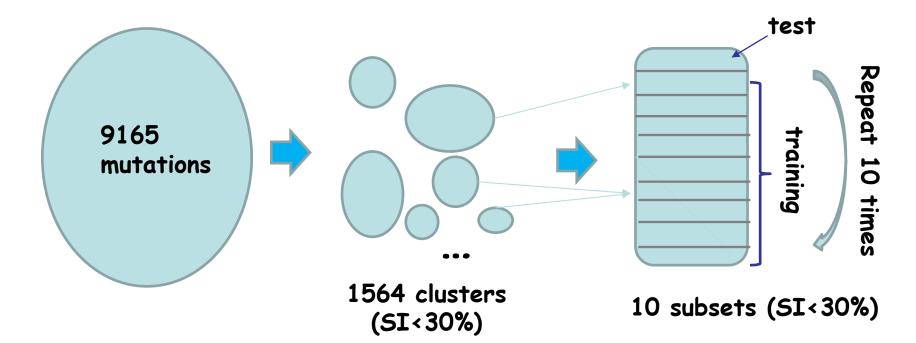
#### Data sets

Disease-associated mutations: 5,356 SNP mutations in 635 proteins

Neural mutations: 3,809 SNP mutations in 1,645 proteins

Total: 9,165 mutations in 1,974 proteins

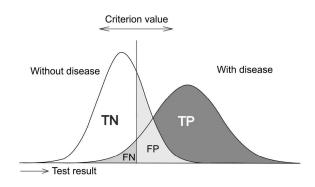
### 10-fold Cross validation procedure



### Benchmark results

#### Results on different training methods

$$\begin{cases} MCC = \frac{TP \times TN - FP \times FN}{\sqrt{(TP + FP)(TP + FN)(TN + FP)(TN + FN)}} \\ ACC = \frac{TP + TN}{TP + FN + TN + FP} \\ Sen(+) = \frac{TP}{TP + FN}, \quad Spe(+) = \frac{TP}{TP + FP} \\ Sen(-) = \frac{TN}{TN + FP}, \quad Spe(-) = \frac{TN}{TN + FN} \end{cases}$$



		<b>Top 20</b> :	features <sup>a</sup>		All features <sup>a</sup>				
	GBC <sup>b</sup>	KNC <sup>b</sup>	SVC <sup>b</sup>	BANN <sup>b</sup>	GBC <sup>b</sup>	KNC <sup>b</sup>	SVC <sup>b</sup>	BANN <sup>b</sup>	
MCC	0.48	0.49	0.50	0.52	0.47	0.44	0.50	0.53	
ACC	0.75	0.75	0.76	0.77	0.74	0.72	0.76	0.77	
SEN <sup>+</sup>	0.81	0.78	0.80	0.82	0.79	0.73	0.78	0.82	
SPE <sup>+</sup>	0.77	0.79	0.79	0.79	0.77	0.79	0.79	0.80	
SEN <sup>-</sup>	0.67	0.71	0.69	0.69	0.68	0.72	0.70	0.71	
SPE <sup>-</sup>	0.71	0.69	0.72	0.74	0.70	0.65	0.71	0.73	

<sup>&</sup>lt;sup>a</sup>Rank of features in S1 Table by the p-value. There are two groups: Top20 and All features. Top 20 are the first 20 lowest features.

BANN is more efficient than other machine-learning methods

<sup>&</sup>lt;sup>b</sup>GBC: gradient boosting classifier; KNC: k-nearest neighbor classifier; SVC: support vector classifier; BANN: Bayes-classifier guided ANN (BANN).

### Benchmark results

#### Comparison of PreDAM with other predictors:

Method	MCC	ACC	Positive		Negative	
			Sensitivity	Specificity	Sensitivity	Specificity
SIFT	0.46	0.74	0.92	0.71	0.49	0.81
SNAP2	0.43	0.75	0.87	0.72	0.52	0.75
PolyPhen2	0.47	0.75	0.89	0.73	0.55	0.78
SNAP2+BANN <sup>a</sup>	0.52	0.77	0.84	0.79	0.69	0.75
PolyPhen2+BANN <sup>b</sup>	0.51	0.76	0.84	0.78	0.67	0.75
PreDAM	0.53	0.77	0.82	0.80	0.71	0.73

<sup>&</sup>lt;sup>a</sup>SNAP2+BANN: the twenty of features extracted from SNAP2 with the lowest p-value. The training method used BANN

- PreDAM output other predictors
- BANN+control > control methods, indicating again BANN is more efficient as machine-learning
- PreDAM > BANN+control, indicating advantage of feature selection in PreDAM

<sup>&</sup>lt;sup>b</sup>PolyPhen2+BANN: the twenty of features extracted from PolyPhen2 with the lowest p-value. The training method used BANN

## Conclusions

- 1. Machine learning is an efficient technique to predict disease-associated SNP mutations
- 2. Bayes-guided neural-network (BANN) training has a higher efficiency than other classifiers and ANN training methods
- 3. Structure based features can improve the accuracy of disease mutation prediction accuracy